

Research article

Effects of hyperbaric pressure on the oxidative stress status in healthy subjects

Efectos de la presión hiperbárica en el estrés oxidativo en sujetos saludables

Ben Huu Nguyen¹ https://orcid.org/0000-0002-4202-6749 Manh Van Phan¹ https://orcid.org/0000-0002-0851-786X Hoan Duy Bui¹ https://orcid.org/0000-0001-6730-4058 Phuong Minh Nguyen¹* https://orcid.org/0000-0003-0948-6871

¹Vietnam Military Medical University. Department of Military Occupational Medicine. Hanoi, Vietnam.

*Author for Correspondence. Email: phuongk21@gmail.com

ABSTRACT

Introduction: In the hyperbaric pressure environment the partial pressure of each gas component increases, which increases oxygen partial pressure. This causes the generation of free radicals and oxidative stress.

Objective: To determine the effects of hyperbaric pressure on the oxidative stress status in healthy subjects.

Methods: 29 healthy men performed standardized hyperbaric chamber dive to a depth of 30 meters of water (msw) for 30 minutes. Blood samples were collected before compression, immediately after decompression and 1 hour after decompression. The levels of Malondialdehyde, Catalase and Superoxide Dismutase were measured in blood samples.

Results: Malondialdehyde activity increased immediately after decompression and recovered at 1 hour after decompression. Superoxide Dismutase enzyme activity decreased immediately after decompression as well as 1 hour after decompression. Catalase enzyme activity increased immediately after decompression, which was significant at 1 hour after decompression.

Conclusion: Changes in the biological markers Malondialdehyde, Catalase and Superoxide Dismutase suggest the appearance of oxidative stress under the influence of a hyperbaric pressure environment. **Keywords:** biological biomarker; catalase; decompression; free radicals; healthy volunteers; malondialdehyde; oxidative stress; superoxide dismutase.

RESUMEN

Introducción: En la condición de presión hiperbárica, la presión parcial de los componentes del aire se encuentra aumentada, incluida la del oxígeno. Esto se considera la causa de formación de radicales libres y el estado de estrés oxidativo.

Objetivo: Determinar los efectos de la presión hiperbárica sobre estado del estrés oxidativo en individuos sanos.

Métodos: 29 hombres sanos realizaron buceo estandarizado en cámara hiperbárica, a una profundidad de 30 metros de agua, durante un tiempo total de 30 minutos. Se recogieron muestras de sangre antes de la compresión, inmediatamente después de la descompresión y una hora después. Se midieron los niveles de malondialdehído, catalasa y superóxido dismutasa en muestras de sangre.

Resultados: La acción del malondialdehído se incrementó inmediatamente después del buceo y se recuperó en 1 hora. La acción de enzima superóxido dismutasa se encontró disminuida al término y 1 hora después, mientras la enzima catalasa se demostró lo contrario y aumentó significativamente en la primera hora.

Conclusión: El cambio de los marcadores biológicos malondialdehído, catalasa y superóxido dismutasa sugiere estado de estrés oxidativo bajo la influencia de presión hiperbárica.

Palabras clave: biomarcadores; catalasa; descompresión; radicales libres; voluntarios saludables; malondialdehído; estrés oxidativo; superóxido dismutasa.

Received: 03/11/2022 Approved: 21/03/2023



INTRODUCTION

In the hyperbaric pressure environment, the partial pressure of each gas component increases, which increases oxygen partial pressure. The increased partial pressure of oxygen leads to the generation of free radicals and oxidative stress. The detrimental effects of exposure to high concentrations of oxygen are due to the increased production of oxygen radicals or other reactive metabolites derived from oxygen.⁽¹⁾ Oxygen is potentially toxic due to the production of free radicals.^(2,3) Oxygen-induced tissue damage is reported for most organs and should be considered when patients are exposed to an elevated oxygen concentration and pressure.^(4,5)

The literature has numerous studies involving hyperbaric pressure in diving. However, these studies were mainly performed on divers,^(6,7) who had many years of dive experience and whose bodies had certain adaptations to the high-pressure environment. Moreover, the previous studies involved mostly wet diving activities; because of this, many other variables can affect the results in addition to the hyperbaric pressure. This includes the diver's underwater activity and the temperature of the aquatic environment. In order to appropriately assess the effects of the hyperbaric pressure environment on oxidative stress status, it is necessary to eliminate the variables introduced by the aquatic environment. It is also necessary to eliminate the variables without prior exposure to the hyperbaric pressure environment. The study's objective was to determine the effects of hyperbaric pressure environment on the oxidative stress status using the biological markers Malondialdehyde, Catalase and Superoxide Dismutase in healthy non-diving subjects.

METHODS

Research design

This study was conducted in the hyperbaric chamber unit (Hyperbarichealth.com, Australia) of the Vietnam Navy Force from April 2020 to May 2020. Data collection and analysis occurred at 3 points in

time: before entering the pressure chamber (before compression), after leaving the pressure chamber (immediately after decompression) and 1 hour after leaving the pressure chamber (1 hour after decompression).

The diving procedure consisted of a dive using compressed air with compression speed of 2 to 2.5 meters/minute, up to a maximum pressure equal to depth of 30 meters of water (msw). When the maximum pressure was reached, decompression was performed immediately and ascent to normal pressure was conducted at 2 to 2.5 meters/minute). Total time in the pressure chamber was 30 minutes (Fig. 1).

Sample collection protocol

Three testing blood samples were collected from the subjects to measure the levels of Malondialdehyde (MDA) and antioxidant enzymes activity of Superoxide Dismutase (SOD) and Catalase (CAT). To exclude the impact of different diets, all subjects were on the same dietary protocol before and during the study.

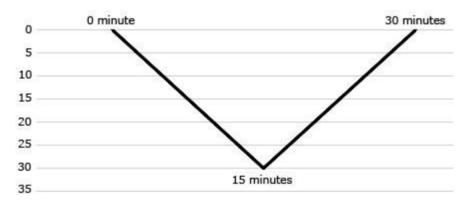


Fig. 1 - Diagram of hyperbaric pressure dive profile.

Biochemical assays

Venous blood samples were collected from the antecubial vein of subjects in proper vacutainers. Blood samples were centrifuged to separate plasma and erythrocytes. Plasma and erythrocytes were immediately stored at -80 °C until use.

Quantification of MDA level: MDA was quantified according to the ELISA method, based on the procedure of ELISA kit (BioVision, Milpitas, CA, USA). Unit of MDA is ng/mL.

Quantification of CAT and SOD: CAT was measured by colorimetry based on the principle that Catalase catalyzes the decomposition of H_2O_2 into H_2O and O_2 ; meanwhile, SOD was quantified by colorimetric method based on the principle that SOD catalyzes the reaction of converting superoxide (O_2^{-}) into O_2 and H_2O_2 ,^(8,9,10) using a kit from BioVision (Milpitas, CA, USA). The CAT and SOD activity unit are mU/mL and ng/mL, respectively.

Subjects

Twenty-nine non-diving healthy men with no acute and chronic medical conditions were recruited to participate in the study. The study was performed in accordance with the procedure of the Vietnamese People's Navy (Vietnam Ministry of National Defense. Circular: Regulations on health standards, prequalification, examination and recruitment, and health assessment of submarine forces of the Navy. Hanoi, Vietnam; 2011).

This protocol had been approved by the ethical committee for biomedical research of Vietnam Military Medical University. All subjects were fully informed about the study process and gave their informed consent.

This study included 29 subjects, with average age of 19.52 ± 1.09 years; average body mass index (BMI) was 19.82 ± 1.32 (kg/m²); average chest round was 83.45 ± 3.11 (cm); the average right-hand force was 39.24 ± 8.74 (kg); the average left-hand force was 36.93 ± 7.74 (kg); average back force was 118.45 ± 20.67 (kg).

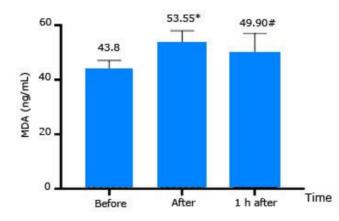
Statistical analysis

The research data was processed by computer using SPSS 22.0 software based on biomedical statistics method. The results were shown in the following form: Mean (\overline{X}), standard deviation (SD), standard error of mean (SEM). Data distribution was checked with the Shapiro – Wilk test and depending on results, the appropriate parametric or non-parametric test was used. Comparison of the means by testing quantitative variables on paired samples (Pair-Sample T Test or Wilcoxon Test). The difference was statistically significant, with p< 0.05.

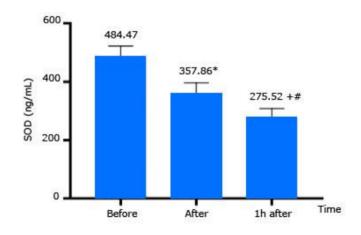


RESULTS

The change of MDA levels is presented in figure 2. MDA activity increased immediately after decompression compared to that before compression. At 1 hour after decompression MDA activity tended to return to the same level as that before compression.



* significant difference between before and after, p= 0.016. $\overline{X} \pm SEM$. # significant difference between after and 1 h after, p= 0.043. $\overline{X} \pm SEM$. **Fig. 2** - Changes of MDA activity after decompression.



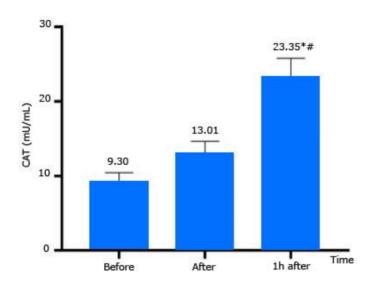
* significant difference between before and after, p = 0.022. X

± SEM.
+ significant difference between before and 1 h after, p < 0.001. X

± SEM.
significant difference between after and 1 h after, p = 0.044. X

± SEM.
Fig. 3 - Changes of SOD activity after decompression.

SOD enzyme activities determined before compression, and immediately after, and 1 hour after decompression are presented in figure 3. SOD enzyme activity decreased significantly after decompression compared to that before compression (p < 0.05).



* significant difference between before and 1 h after, p < 0.001. X ± SEM.
significant difference between after and 1 h after, p < 0.001. X ± SEM.
Fig. 4 - Changes of CAT activity after decompression.

CAT enzyme activities determined before compression, and immediately after, and 1 h after decompression are presented in figure 4. CAT enzyme activity at 1 h after decompression increased more than that before compression, and immediately after decompression significantly (p < 0.05).

DISCUSSION

MDA activity: In this study MDA increased immediately after decompression and decreased gradually at one hour after decompression (Fig. 2). *Ferrer* et al.⁽⁵⁾ studied the effects of high pressure on a group of divers performing a dive at a depth of 40 meters breathing compressed air with a total time of 25 minutes,

versus another group of healthy males breathing oxygen at a pressure of 2.2 ATA for 60 minutes in a pressure chamber. The results showed that MDA activity increased after hyperbaric exposure, but it was not statistically significant. The study by *Sureda* et al.⁽¹¹⁾ was performed on 7 professional divers conducting a dive at a maximum depth of 40 meters for total time of 25 minutes and breathing compressed air. The ascent process took a 5-minute safety stop at the 3-meter decompression station. After the dive, it was reported that MDA activity had some statistically significant changes.

Kozakiewicz et al.⁽¹²⁾ compared diver's MDA activity before and after exposure to hyperbaric pressure in a pressure chamber equivalent to 30 msw (4 ATA). The result was that erythrocyte MDA activity increased immediately after being exposed to hyperbaric pressure; the change was statistically significant. In 2012, *Sureda* et al.⁽⁶⁾ studied 9 professional divers breathing compressed air diving at a depth of 50 meters for a total time of 35 minutes. The study demonstrated that MDA did not increase immediately after diving, but 3 hours after diving MDA activity increased significantly compared to before diving.

The results of this study showed an earlier increase in MDA compared to *Sureda*'s et al.⁽⁶⁾ study, and the MDA activity in the current study was lower. We believe this is due to the fact that the in current study subjects were healthy young men who had never participated in diving. The MDA activity in the current study ranged from 40 to 60 ng/mL (0.56-0.83) μ mol/L. MDA molecular weight (72.07) is equivalent to healthy human MDA activity.⁽¹³⁾ However, subjects in the *Sureda*'s et al.⁽⁶⁾ study were professional divers, so these subjects had adapted to oxidative stress, with high MDA activity.

SOD activity: Results in this study demonstrated that SOD activity decreased after decompression (Fig. 3), which was not consistent with some other studies. The study by *Sureda* et al.⁽¹¹⁾ was conducted with dives to a maximum depth of 40 meters, for a total time of 25 minutes breathing compressed air. The results of this study showed that, although plasma SOD activity increased immediately but not significantly after the dive, up to 3 hours after the dive, plasma SOD activity increased more significantly than before the dive. The activity of antioxidant enzyme in red blood cells did not have a statistically significant change. In 2012, *Sureda* et al.⁽⁶⁾ reported an increase in activity of SOD enzymes after the dive to a depth of 50 meters, for a total time of 35 minutes while breathing compressed air. *Radojevic-Popovic* et al.⁽¹⁴⁾ studied 32 professional divers with an average age of 32 ± 5.1 years and diving experience

from 3 to 18 years. They made dives to 30 meters' depth breathing compressed air for 30 minutes. Their descent speed was 10 meters/minute, the ascent speed was 9 meters/minute, with a 3-minute safety stop at 3 meters. The results demonstrated that SOD activity decreased after diving (before diving 1 416.26 \pm 334.15 U/g Hb x 10³; immediately after diving: 1 275.91 \pm 378.51 U/g Hb x 10³), however, this change was not statistically significant.

Adaptation of the antioxidant system that occurred in professional divers was considered to be an important factor in the absence of significant changes in the activity of SOD. A study by *Zwart* et al.⁽¹⁵⁾ evaluated the effects of a long-term dive on SOD change and found the decrease in SOD activity after the dive. Based on results in current study and others, professional divers adapted to hyperbaric pressure environment, so the change in this enzyme activity seems to be of slower appearance. In addition to the long period of the dive, can be a reason which led to the presence of oxidative stress, and the antioxidant capacity of SOD enzyme can be decreased. Subjects in current study are healthy men, who do not adapt to high pressure environment, their response can occur earlier and stronger than the that of professional divers.

CAT activity: Catalase catalyzes the decomposition of H_2O_2 to water and oxygen.^(8,9,10) Change of CAT activity suggest that oxidative stress status appeared. In this study, CAT increased immediately after decompression, which was significant at 1 hour after decompression (Fig. 4) and consistent with other studies. *Ferrer* et al.⁽⁵⁾ studied the effects of high pressure on the antioxidant system of lymphocytes on a group of 7 divers with a mean age of 26.0 ± 4.7 years old. They performed a dive to 40 meters breathing compressed air for a total time of 25 minutes. This included 10 minutes at 40 meters depth and 5 minutes at a 3-meter decompression station. The results of this group showed that CAT activity increased significantly at 3 hours post-dive ($27.9 \pm 2.2 \text{ K}/10^9$ lymphocytes) compared to before dive ($21.4 \pm 1.1 \text{ K}/10^9$ lymphocytes). Another group of this study consisted of 12 healthy males with mean age of 25.3 ± 3.9 years, breathing oxygen at a pressure of 2.2 ATA for 60 minutes in a pressure chamber. The results showed an increase in the activity of CAT after the end of hyperbaric oxygen pressure breathing ($26.2 \pm 2.2 \text{ K}/10^9$ lymphocytes) compared to before exposure to hyperbaric oxygen $22.2 \pm 2.4 \text{ K}/10^9$ lymphocytes), but the difference was not significant.⁽⁵⁾

Sureda et al.⁽¹¹⁾ conducted research on diving to a maximum depth of 40 meters with a total time of 25 minutes while breathing compressed air. The results of this study showed that plasma CAT activity

increased significantly immediately after the dive $(276 \pm 23 \text{ Kat/l})$ compared with before the dive (200 \pm 18 Kat/l); 3 hours after completion of the dive, CAT activity recovered, which was similar to before diving. Meanwhile, the activity of this antioxidant enzyme in red blood cells did not have a statistically significant change. In 2012 *Sureda* et al.⁽⁶⁾ conducted a research on diving to a depth of 50 meters with in a total time 35 minutes while breathing compressed air; they reported an increase in the activity of CAT enzyme after the dive.

Gasier et al.⁽⁷⁾ performed a study on 100% oxygen breathing for 2 hours at 1 ATA, 1.5 ATA and 2 ATA in the pressure chamber. The study showed that erythrocyte CAT enzyme activity 1 hour after breathing oxygen was significantly reduced compared to before breathing oxygen (1.5 ATA); at 2 ATA pressure it was observed to decrease at all 3 time points, after 15 minutes, after 1 hour, and after 2 hours breathing oxygen compared to before breathing oxygen. Meanwhile, plasma CAT activity did not change significantly following all of the hyperbaric oxygen exposures.⁽⁷⁾ *Radojevic-Popovic* et al.⁽¹⁴⁾ performed diving with compressed air to 30 meters for 30 minutes. The results showed that there was an increase in CAT activity immediately after diving (before diving 5.21 ± 2.38 U/g Hb x 10^3 ; immediately after diving 6.39 ± 3.25 U/g Hb x 10^3); but, this change was not significant. The authors reasoned that the study subjects were professional divers with frequent diving, so the antioxidant system had adjusted already in the direction of adaptation. Therefore, a new dive did not lead to any changes in CAT activity.

The results of this study have some similarities but also some differences compared with the results of other authors. In author's opinion this is due to the following factors: First, during dry diving in the pressure chamber the subjects did not actually carry out any physical activities. This is different from these previous studies, where subjects conducted dives with different activities to perform descent and ascent. Second, our study subjects were healthy men exposed to a hyperbaric barometric pressure environment for the first time, so they had not yet adapted to these environmental conditions. Thus, changes of oxidative stress (MDA activity) occurred more rapidly and antioxidants (SOD enzyme and CAT enzyme activities) changes early, which is different from previous studies.

As a limitation, the current study followed the impact of a hyperbaric pressure environment in a short duration of only 1 hour after leaving the pressure chamber, lacking a longer follow-up time of 3 to 6

hours after hyperbaric pressure. Longer follow up will yield more information on the evolution, resilience as well as the changeable level of biologic markers.

In conclusion, some changes in the biologic markers MDA, CAT and SOD suggest the appearance of oxidative stress under the influence of a hyperbaric pressure environment.

Acknowledgement

We are grateful to our subjects for their participation this research. Also, we would like to thank hyperbaric chamber unit of the Vietnam Navy Force for their technical assistance in this study.

BIBLIOGRAPHIC REFERENCES

 Narkowiczt CK, Vial JH, Mccartney PW. Hyperbaric oxygen therapy increases free radical levels in the blood of humans. Free Radic Res Commun. 1993; 19(2):71-80. DOI: 10.3109/10715769309056501
 Bin-Jaliah I, Dallak M, Haffor ASA. Effect of hyperoxia on the ultrastructural pathology of alveolar epithelium in relation to glutathione peroxidase, lactate dehydrogenase activities, and free radical production in rats, rattus norvigicus. Ultrastruct Pathol. 2009; 33:112–22. DOI: 10.1080/01913120902889179

 Jamieson D, Chance B, Cadenas E, Boveris A. The relation of free radical production to hyperoxia. Annu Rev Physiol. 1986; 48:703-19. DOI: 10.1146/annurev.ph.48.030186.003415
 Pott F, Westergaard P, Mortensen J, Jansen EC. Hyperbaric oxygen treament and pulmonary function. Undersea Hyperb Med. 1999 [access: 01/09/1999]; 26(4):225-28. Available from:

https://pubmed.ncbi.nlm.nih.gov/10642068

5. Ferrer MD, Sureda A, Batle JM, Tauler P, Tur JA, Pons A. Scuba diving enhances endogenous antioxidant defenses in lymphocytes and neutrophils. Free Radic Res. 2007; 41(3): 274–281. DOI: 10.1080/10715760601080371

6. Sureda A, Batle JM., Ferrer MD, Mestre-Alfaro A, Tur JA, Pons A. Scuba diving activates vascular antioxidant system. Int J Sports Med. 2012; 33:531–6. DOI: 10.1055/s-0031-1297957

7. Gasier HG, Fothergill DM. Oxidative stress, antioxidant defenses and nitric oxide production following hyperoxic exposures. Undersea Hyperb Med. 2013 [access: 21/05/2013]; 40(2):125-34. Available from: https://pubmed.ncbi.nlm.nih.gov/23682544/

8. Kasperczyk S, Birkner E, Kasperczyk A, Zalejska-Fiolka J. Activity of superoxide dismutase and catalase in people protractedly exposed to lead compounds. Ann Agric Environ Med. 2004 [access: 11/10/2004]; 11: 291 - 296. Available from:

https://www.proquest.com/openview/10c7238e1d4f603cf07810938d672190/1?pqorigsite=gscholar&cbl=5221234

9. Laher I. Systems biology of free radicals and antioxidants. Vancouver, British Columbia, Canada: Springer; 2014.

10. Bhattacharyya A, Chattopadhyay R, Mitra S, Crowe SE. Oxidative stress: an essential factor in the pathogenesis of gastrointestinal mucosal diseases. Physiol Rev. 2014; 94:329 - 354. DOI:

10.1152/physrev.00040.2012

11. Sureda A, Ferrer MD, Batle JM, Tauler P, Tur JA, Pons A. Scuba diving increases erythrocyte and plasma antioxidant defenses and spares NO without oxidative damage. Med Sci Sports Exerc. 2009; 41(6):1271-6. DOI: 10.1249/MSS.0b013e3181951069

12. Kozakiewicz M, Kędziora-Kornatowska K, Kaczerska D, Siermontowski P, Olszanski R, Krefft K. Influence of exposure in hyperbaric chambers on selected parameters of oxidative stress in professional divers. Undersea Hyperb Med. 2018 [access: 01/01/2018]; 45(1):49-54.

https://europepmc.org/article/med/29571232

13. Bhutia Y, Ghosh A, Sherpa ML, Pal R, Mohanta PK. Serum malondialdehyde level: surrogate stress marker in the sikkimese diabetics. J Nat Sci Biol Med. 2011; 2(1):107-12. DOI: 10.4103/0976-9668.82309

14. Radojevic-Popovic R, Zivkovic V, Jeremic N, Sretenovic J, Velicanin N, Bradic J, et al. An evaluation of the redox state in professional scuba divers. Undersea Hyperb Med. 2015 [access: 01/09/2015]; 42(5):409-16. https://pubmed.ncbi.nlm.nih.gov/26591980/



15. Zwart SR., Kala G, Smith SM. Body iron stores and oxidative damage in humans increased during and after a 10- to 12-day undersea dive. J Nutr. 2008; 139:90–5. DOI: 10.3945/jn.108.097592

Conflict of interest

The authors have no conflicts of interest to disclose.

Authorship contribution

Conceptualization: Phuong Minh Nguyen, Ben Huu Nguyen.
Data curation: Ben Huu Nguyen, Manh Van Phan.
Formal analysis: Ben Huu Nguyen, Hoan Duy Bui.
Research: Phuong Minh Nguyen, Ben Huu Nguyen, Manh Phan Van, Hoan Duy Bui.
Methodology: Phuong Minh Nguyen, Ben Huu Nguyen.
Supervision: Phuong Minh Nguyen.
Validation: Phuong Minh Nguyen.
Display: Phuong Minh Nguyen, Ben Huu Nguyen.
Drafting - original draft: Phuong Minh Nguyen, Ben Huu Nguyen.
Drafting - revision and editing: Phuong Minh Nguyen, Ben Huu Nguyen, Ben Huu Nguyen.