

STANDARD TEST OF LABELLING EFFICIENCY FOR QUALITY CONTROL OF NO CARRIER ADDED $^{90}\text{YCl}_3$

Milos Beran, K. Eigner Henke, J. Srank, F. Melichar
Department of Radiopharmaceuticals
The Institute of Nuclear Physics, The Academy of Sciences of the Czech Republic (INP ASCR)
Rez near Prague, Czech Republic
milber@attas.cz, beran@ujf.cas.cz

Abstract

Particle emitting radionuclides (e.g. β -emitters ^{90}Y and ^{177}Lu , β -emitter ^{149}Tb , Auger electron emitter ^{165}Er or positron emitter ^{88}Y) are more and more frequently used in research and clinical practice for imaging and radionuclide targeted therapy in nuclear medicine. These radiometals, altogether trivalent lanthanides or actinides with high specific radioactivity, coupled to biomolecule carriers (peptides or monoclonal antibodies) through chelating link (e.g. DTPA or DOTA) bind to specific antigens and/or receptors of diseased tissues, which enables the imaging (positron emitters) or destruction (β^- , α^- , and Auger electron emitter) of the diseased tissue releasing the antigens or carrying the receptors. The radionuclide precursor $^{90}\text{YCl}_3$ (solution of hard β^- -emitter ^{90}Y in diluted HCl) with high purity and specific activity is already commercially produced and successfully used in nuclear medicine, e.g. for radioimmunotherapy (RIT) of Lymphoma. Specification and purity of our product obtained using extraction $^{90}\text{Sr}/^{90}\text{Y}$ generator (using technology of centrifuge extractors with di-2-ethylhexylphosphoric acid, D2EHPA) is examined and compared to other similar products in this contribution. A standard method for determination of labelling efficiency of the $^{90}\text{YCl}_3$ precursor based on its reaction with DOTA-Tyr³-Octreotide (DOTA-TOC) and ITLC-SG chromatographic separation is described and proposed for the quality control.

PRUEBA ESTÁNDAR DE LA EFICIENCIA DEL MARCAJE PARA EL CONTROL DE LA CALIDAD DEL $^{90}\text{YCl}_3$ SIN PORTADOR AÑADIDO

Resumen

Los radionucleidos emisores de partículas (ej. emisores β^- ^{90}Y y ^{177}Lu , emisores α^- ^{149}Tb , emisor de electrones Auger ^{165}Er o emisor de positrones ^{88}Y) se emplean cada vez más en la investigación, en la práctica clínica para el procesamiento de imágenes y en la terapia dirigida de radionucleidos en medicina nuclear. Estos radiometales junto con actínidos o lantánidos trivalentes con alta actividad específica, asociados a portadores de biomoléculas (péptidos o anticuerpos monoclonales) por medio de un enlace con quelatos (ej. DTPA o DOTA) se unen a antígenos específicos receptores de tejidos afectados que permiten el procesamiento de imágenes (emisores de positrones) o la destrucción (β^- , α^- , y emisores de electrones de Auger) de los tejidos afectados que liberan los antígenos o portan los receptores. El precursor del radionucleido $^{90}\text{YCl}_3$ (solución de emisor β^- duro - ^{90}Y en HCl diluido) con elevada pureza y actividad específica ya se produce comercialmente y se usa exitosamente en la medicina nuclear, por ejemplo, para la radioimmunoterapia (RIT) del linfoma. En esta contribución se examina la especificación y la pureza de nuestro producto, obtenido mediante extracción con generador $^{90}\text{Sr}/^{90}\text{Y}$ (empleando la tecnología de extractores por centrifugado con ácido di-2-ethylhexyl phosphoric D2EHPA) y se compara con otros productos similares. Se describe y propone para el control de la calidad un método estándar para determinar la eficiencia en el marcaje del precursor del $^{90}\text{YCl}_3$ basado en su reacción con DOTA-Tyr³-ostreotido (DOTA-TOC) y separación cromatográfica ITLC-SG.

Key words: fluorine 18, fluorodeoxyglucose, liquid column chromatography, positron computed tomography, quality control, radiopharmaceuticals, toxicity, autoradiolysis

INTRODUCTION

The internal targeted radionuclide therapy for treatment of various diseases in human oncology has recently reached the age of mature. In February 2002, ^{90}Y -ibritumomab tiuxetan (Zevalin; IDEC Pharmaceuticals Corp., San Diego, CA) monoclonal antibody (mAb) received the final approval by the

Food and Drug Administration (FDA) as the first commercially available radiolabelled antibody for cancer treatment. Schering received the European approval for Zevalin on January 22, 2004. Zevalin proved high efficiency in treatment of relapsed or refractory non-Hodgkin's *Lymphoma* of B-cells in clinical trials [1]. An overall response rate of 80% and a complete response of 30% were observed

compared to the 56% and 16% response for immunotherapy with Rituximab, unlabelled mAb alone. Since that time, very intensive research of various biomolecules labelled with different therapeutic radionuclides, targeted against the most significant tumour antigens and receptors, has started. For diagnostics of chosen types of tumors, radioactive labelled octreotide derivatives are often indicated. Octreotide is a synthetic somatostatin analogue with a longer biological half-life. It is metabolised in the liver. Much like somatostatin, octreotide binds to the SRIF 1-class SSRT (SRIF: somatotropine release inhibiting factor). ^{90}Y -DOTA-d-Phe¹-Tyr³-octreotide (^{90}Y -DOTATOC) has recently been used for treatment of patients with somatostatin receptor positive tumors. A pilot study with 44 patients with advanced somatostatin receptor positive tumors showed that only 29 of the 44 patients could be treated with ^{90}Y -DOTATOC. Taking into account the radiotoxicity of ^{90}Y , a reliable diagnostic tool for examination of the ^{90}Y -DOTATOC affinity to the somatostatin receptor positive tumor is essential to avoid the "try and error principal" therapy. Patients with a cumulative radiation dose $\leq 7400 \text{ MBq/m}^2$ showed no severe renal or hematological toxicity, while the patients having received a cumulative radiation dose $> 7400 \text{ MBq/m}^2$ developed renal and/or hematological toxicity [2]. Therefore, to develop and optimize a new methodology for synthesis of $^{86/90}\text{Y}$ -DOTATOC derivatives appears to be very useful. The biochemical and biophysiological properties of the octreotide derivatives are not influenced by the used yttrium isotope.

Very high demands are imposed on purity of radionuclide precursors used for labelling of biomolecules in nuclear medicine. There are only few β -emitting radionuclides commercially produced in corresponding quality. Among them, no carrier added $^{90}\text{YCl}_3$ radionuclide precursor in diluted $0.04 - 0.05 \text{ mol.l}^{-1}$ hydrochloric acid is currently the most important because of the very high ^{90}Y specific radioactivity (theoretically about 500 Ci.mg^{-1}) being reached in the production and the ^{90}Y decay properties (pure β -emission; average energy 0.935 MeV , maximum 2.284 MeV ; penetration in tissue $5-9 \text{ mm}$; physical half-life 64.1 h).

Some of the commercially produced and marketed ^{90}Y radionuclide preparations, including their main properties as stated in producer's specifications, are presented in table 1.

The quality of the latest $^{90}\text{YCl}_3$ commercial product approved in Europe as radiopharmaceutical precursor (Yttriga, AEA Technology, QSA Global GmbH, Braunschweig, Germany) is estimated using the binding efficiency test with DTPA instead of chemical impurities definition. This test seems to be much more predicative and crucial for quality control of preparations than estimating the admissible limits of individual chemical impurities. Determination of all trace impurities and estimation of their acceptable concentrations are nearly impossible. Therefore, a Standard Labelling Test (SLT) using DOTATOC, one of the DOTA conjugated peptides frequently used in RIT preclinical and clinical experiments, has been developed in our laboratory. The results are presented in this contribution.

Experimental

High pure $^{90}\text{YCl}_3$ radionuclide precursor was prepared in our laboratory using the experimental pilot facility with two-stage Centrifugal Semicounterflow Extractors [3]. The separation of ^{90}Y from the $^{90}\text{Sr}/^{90}\text{Y}$ mixture in 0.5 mol.l^{-1} nitric acid was accomplished by extracting it with di-2-ethylhexylphosphoric acid in n-dodecane. The extraction was followed by washing steps (e.g. with n-hexane) and reextraction of ^{90}Y into 5 mol.l^{-1} hydrochloric acid. Vacuum evaporation was used for the final concentration. Subsequently, the product was dissolved in a small volume of $0.05 \text{ mol.l}^{-1} \text{ HCl}$. The characteristics of individual production batches used in development of Standard Labelling Test are summarized in table 2. The two last batches significantly contaminated by Fe were used to determine the level of Fe resulting in rapid decrease of ^{90}Y labelling efficiency.

The main quality parameters of our preparation in comparison to the Yttriga product (AEA Technology, QSA Global GmbH, Braunschweig, Germany) are presented in table 3.

DOTA-Tyr³-Octreotide, DOTA-TOC (MW $1421.64 \text{ g.mol}^{-1}$), was supplied by piCHEM, Austria (white lyophilisate, purity $> 95\%$ by HPLC).

Labelling Protocol for analytical use [4]: About $2.5-5 \text{ MBq}$ of $^{90}\text{YCl}_3$ in $300 \mu\text{l}$ of $0.05 \text{ mol.l}^{-1} \text{ HCl}$ was mixed with $5-20 \mu\text{g}$ of DOTA-TOC in $300 \mu\text{l}$ of 0.4 mol.l^{-1} sodium acetate (NaAc – supra pure) and

Table 1. Commercial products survey of no carrier added $^{90}\text{YCl}_3$ radionuclide precursors

Product (Company)	RCH Purity ($^{90}\text{YCl}_3$,%)	^{86}Sr impurity (kBq-GBq ⁻¹ of ^{90}Y)	Main chemical impurities
Yttricia (CIS)	≥ 97.0	≤ 20	Fe $\leq 10 \mu\text{g.ml}^{-1}$
Yttrium-90 (Perkin-Elmer)		< 2.5	Fe $< 20 \mu\text{g.Ci}^{-1}$
Yttriga (QSA Global GmbH)	> 99	< 10	DTPA-binding Test $> 80\%$
Yttrium-90 (MDS Nordion) - Canada - Belgium	≥ 95	≤ 20 ≤ 2.5	(Fe+Zn+Cu+Cd+Pb) $\leq 30 \mu\text{g.Ci}^{-1}$

Table 2. Summary of individual $^{90}\text{YCl}_3$ production batches used for development of SLT

Batch N°	Volume activity (GBq.ml ⁻¹)	RCH Purity ($^{90}\text{YCl}_3$,%)	^{90}Sr impurity (kBq.GBq ⁻¹ of ^{90}Y)	Fe impurity (µg.ml ⁻¹)
060807-075zk	0.97	98.3	1.81	1.04
060814-076zk	0.64	99.9	1.53	1.23
060821-077zk	1.42	99.9	0.99	1.54
060904-079zk	1.23	99.6	0.98	0.55
060925-080zk	1.58	99.4		40.8
061002-081zk	1.45	99.5		36.3

Table 3. Main quality parameters of [^{90}Y]yttrium chloride produced in INP ASCR and Yttriga

Batch N°	RD (YYYY/MM/DD-hh:mm)	Volume activity (GBq.ml ⁻¹)	RCH Purity ($^{90}\text{YCl}_3$,%)	Fe impurity (µg.ml ⁻¹)
083zk	2006/10/18-13:30	0.5	99.8	2.39
086zk	2006/11/06-16:30	1.08	99.8	1.79
Yttriga	2006/11/07-12:00	2.0	99.9	2.61

heated in water bath for 3-40 min at 60-90 °C in order to find the optimal labelling conditions.

Chromatographic method for determination of ^{90}Y fraction incorporated in DOTA-TOC complex: About 2 µl of the solution after termination of Labelling Protocol was applied on the start (about 10 mm at the margin) of ITLC chromatographic strips (15 x 200 mm, ITLC-SG, Pall Life Sciences, PALL Corporation). Subsequently, the strips were ascendingly developed in 0.1 mol.l⁻¹ sodium citrate (Na₃Cit) up to about 10 mm below the strip margin. The strips were analyzed on ^{90}Y distribution using β-scanner after being air dried. Each sample was analyzed in 3 parallels.

Instruments: Linear scanning of radiochromatograms for ^{90}Y distribution was done by TLC Scanner RAYTEST, Minigita with Proportional Gas Flow (Argon 90%-Methane 10%) Beta Counting Tube (Isotopenmeßgeräte GmbH).

RESULTS AND DISCUSSION

Bifunctional chelators such as 1,4,7,10-tetraazacyclododecane-N,N',N'',N'''-tetraacetic acid (DOTA) and diethylenetriaminepenta-acetic acid (DTPA) [5] are widely used to bound radiometals, namely trivalent lanthanides and actinides, onto biomolecules. The DOTA bioconjugates demonstrate much higher *in vivo* stability than the DTPA analogues. However, the kinetics of their synthesis (e.g. chelation of ^{90}Y by DOTA) is significantly slower and requires temperature range up to 100°C. Nevertheless, they are preferred for use in nuclear medicine, and,

thereby, they represent suitable examples for developing SLT. Such test of labelling efficiency reflects credibly the labelling power of $^{90}\text{YCl}_3$ radionuclide precursor in majority of its applications. Very high attainable labelling efficiency despite the presence of chemical impurities (e.g. metal interferents like iron) in low concentrations is an added reason for choosing the ^{90}Y -DOTATOC system for SLT. Figure. 1 demonstrates the typical chromatogram appearance of SLT (Batch No. 060904-079zk of our production - table 2).

Separation of ^{90}Y -DOTATOC complex from free yttrium-90 is quantitative in the presented chromatographic system. These only two components appear in solutions after the labelling procedure described above so that analytical evaluation of results is very easy. More precise determination of optimal labelling

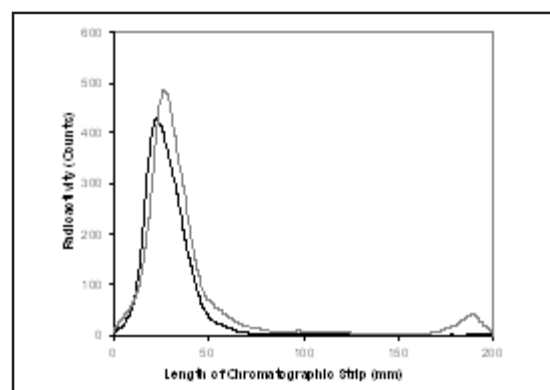


Figure. 1. Typical Chromatograms of Y-90 Labelled DOTA-TOC.

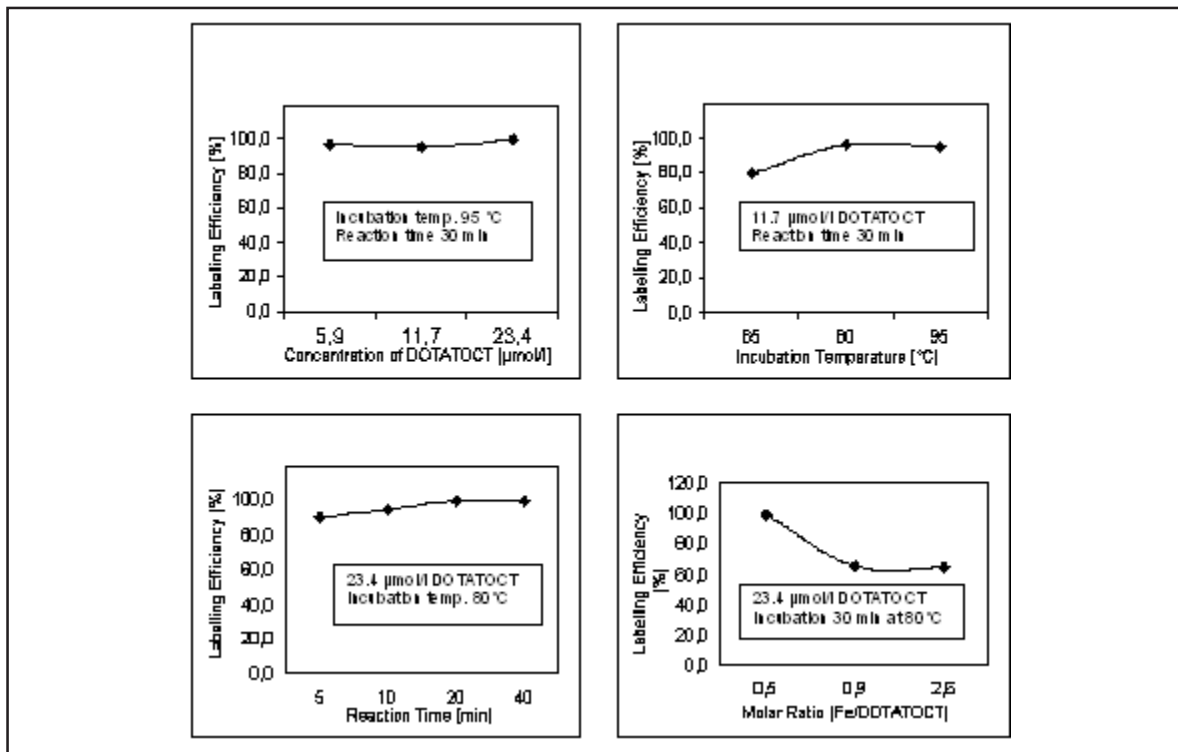


Figure 2. Dependence on DOTATOCT concentration. Figure 3. Dependence on reaction temperature. Figure 4. Reaction rate. Figure 5. Dependence on molar ratio of iron to DOTATOCT.

conditions (labelling kinetics, incubation temperature, concentration of DOTATOCT) is illustrated in figures 2-4. Obviously, the optimal incubation temperature (maximum yield) is 80°C. Higher temperature may cause degradation of the peptide resulting in decreased labelling yields. The optimal reaction time seems to be 25-30 minutes because of slower formation kinetics of DOTA complexes.

The Standard Labelling Protocol was precised as follows:

About 2.5–5 MBq of $^{90}\text{YCl}_3$ in 300 µl of 0.05 mol.l⁻¹ HCl is mixed with 20 µg of DOTATOCT in 300 µl of 0.4 mol.l⁻¹ sodium acetate (final concentration of DOTATOCT 23.4 µmol/l) and heated in water bath for 30 min at 80°C. ITLC chromatographic procedure (see page 4) is then applied (after cooling down the sample to the room temperature) to determine $^{90}\text{YCl}_3$ labelling efficiency. Each chromatogram is developed in 3 parallels. Two production batches of $^{90}\text{YCl}_3$ radionuclide precursor with higher Fe content (080zk and 081zk – see table 2) were analyzed using this method (SLT) in order to determine maximum permissible limit for Fe impurity. This limit corresponds to stoichiometric saturation of DOTATOCT (figure 5). Acceptable labelling yield was obtained up to molar ratio $[\text{Fe}/\text{DOTATOCT}] = 0.5$.

The SLT developed in our laboratory and described above was used to estimate the labelling efficiency of two $^{90}\text{YCl}_3$ radionuclide precursor products compiled in table 3:

Yttrigra-06/11/07, QSA Global GmbH and our product Batch No. 086zk-06/11/06. Labelling efficiency (mean ± SD, n=3) for both products was: 99.5 ± 0.1% for Yttrigra and 96.1 ± 0.8% for our product.

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